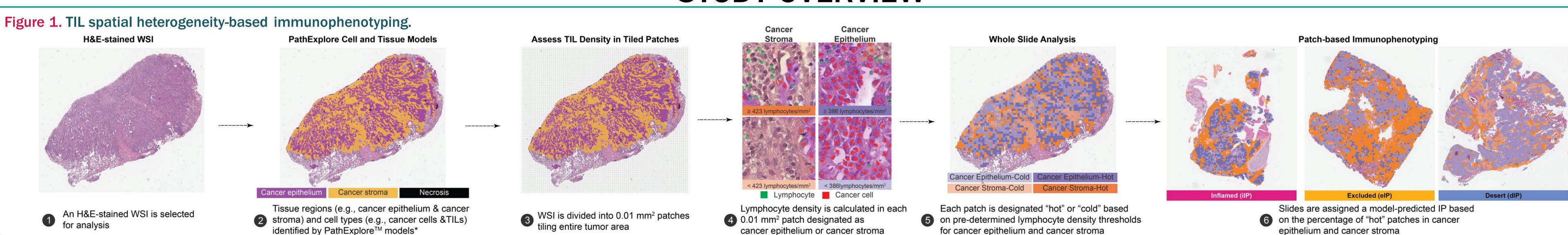
Abstract #8539

Nhat Le^{1*}, Laura Dillon^{2*}, Ciyue Shen¹, Matthew Bronnimann¹, Jun Zhang¹, Jennifer Hipp¹, Tan Nguyen¹, Michael Griffin¹, Michael Griffin¹, Michael Griffin¹, Michael Griffin¹, Michael Griffin¹, Michael Hercessian¹, Jennifer Hipp¹, Tan Nguyen¹, Michael Griffin¹, Michael Griffi Xinwei Sher², G. Travis Clifton^{2*}, <u>Bahar Rahsepar</u>^{1*}

STUDY BACKGROUND

- The classification of tumors as inflamed, excluded or desert based on spatial patterns of tumor infiltrating lymphocytes (TILs)¹ is a potential biomarker of patients likely to respond to checkpoint inhibitors (CPI)². However, the subjectivity of manual methods to assess these immune phenotypes (IPs) and poor standardization in the methods and thresholds to define IPs have hampered their clinical adoption^{3,4}.
- Here, we describe a data-driven approach to inform IP threshold selection based on predicted lymphocyte densities in patches of hematoxylin and eosin (H&E)-stained whole slide images (WSI) by maximizing differences in overall survival (OS) between IPs.

STUDY OVERVIEW



METHODS

- H&E-stained WSI (N=4,082) from multiple datasets from the cancer genome atlas (TCGA; COAD, READ, SKCM, PRAD, ESCA, STAD, PAAD, thresholds for IPs.
- Two cohorts of patients with NSCLC were used to assess the clinical

Immune phenotype prediction

¹PathAl, Inc., Boston, MA

CONTACT

Bahar Rahsepar: bahar.rahsepar@pathai.com Laura Dillon: laura.dillon@incendiatx.com

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diagnostic procedures.





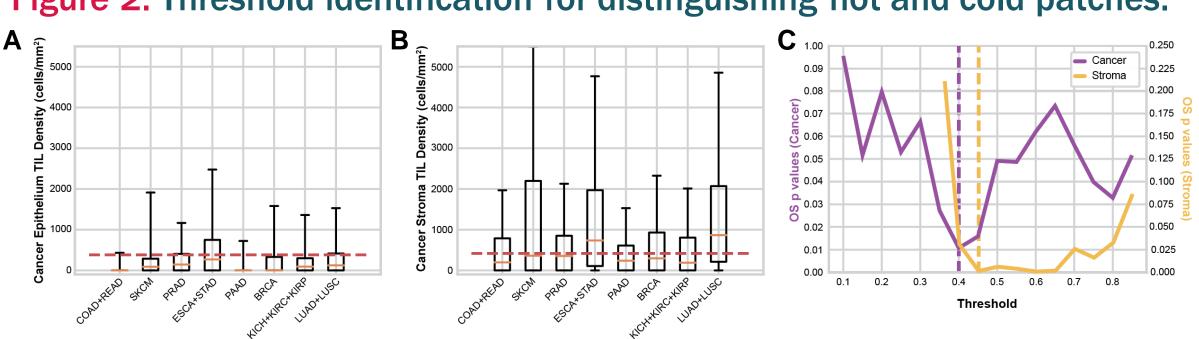


BRCA, KICH, KIRC, KIRP, LUAD, LUSC)⁵ were used to determine

implications of IPs predicted by our approach: 1) TCGA cohort, consisting of LUAD (N=459) and LUSC (N=424) and 2) a clinical cohort consisting of PD-(L)1 inhibitor-treated NSCLC patients (N=95) enrolled in the BIP precision medicine study (NCT02534649; Institut Bergonié, Bordeaux, France).

- A model to classify the IPs of NSCLC samples from H&E images was developed using PathExplore⁶ models as described in Fig. 1.
- Lymphocyte densities were extracted for 0.01 mm² patches tiled across WSI. Cut-offs to define cancer epithelium and cancer stroma patches as hot or cold were defined based on the 75th and 50th percentiles, respectively, of lymphocyte densities in cancer epithelium and cancer stroma (Fig. 2A,B).
- Hierarchical fitting yielded optimal thresholds in cancer epithelium and cancer stroma (Fig. 2C) that minimize p-values of OS differences between IPs.

Figure 2. Threshold identification for distinguishing hot and cold patches.



Selected lymphocyte density thresholds in (A) cancer epithelium (75th percentile across all sampled patches from all indications) and (B) cancer stroma (50th percentile across all sampled patches from all indications) for distinguishing hot and cold patches. C) Thresholds (dashed lines) were selected to minimize the p-values of OS differences between IPs.

Exploratory Analyses

- Model-predicted IPs were compared to progression-free survival (PFS) and overall survival (OS) in both the TCGA and clinical cohorts. False discovery rate (FDR) correction was done with Benjamini-Hochberg.
- Survival was also assessed in the clinical cohort using PD-L1 tumor proportion score (TPS), iIP status, and TIL density as covariates.

Table 1. Thresholds chosen for IP prediction in NSCLC.

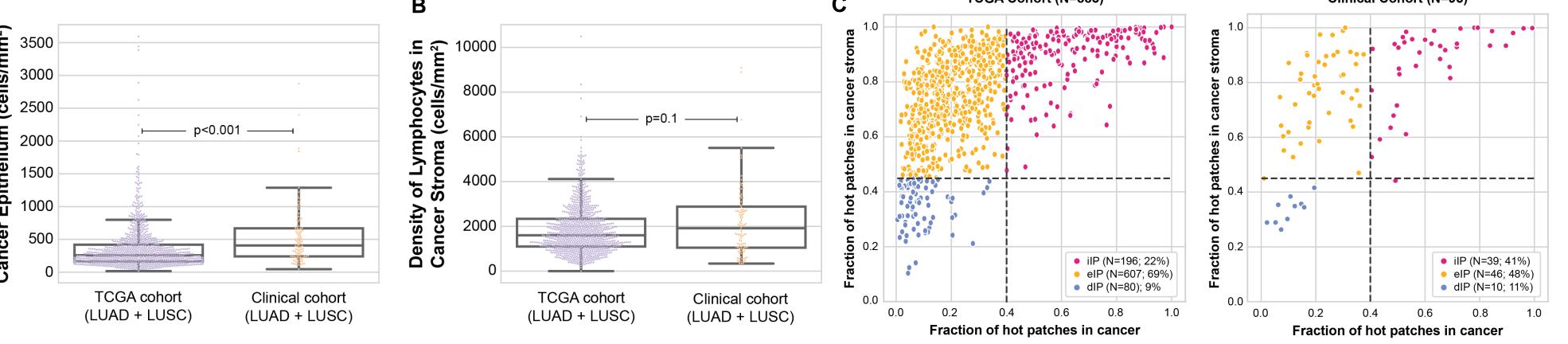
	Model Predicted IP	Criteria
	Inflamed (iIP)	>40% hot patches in cancer epithelium
	Excluded (eIP)	≤40% hot patches in cancer epithelium; >45% hot patches in cancer stroma
	Desert (dIP)	≤40% hot patches in cancer epithelium; ≤45% hot patches in cancer stroma
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In the TCGA NSCLC cohort, model-predicted iIP and eIP patients had significantly better OS compared to dIP (HR=0.53, p=0.003 and HR=0.59, p=0.003, respectively; Fig. 4A). In the clinical cohort, PFS was significantly shorter in model-predicted eIP patients compared to iIP (HR=0.54, p=0.045; Fig. 4B).

RESULTS

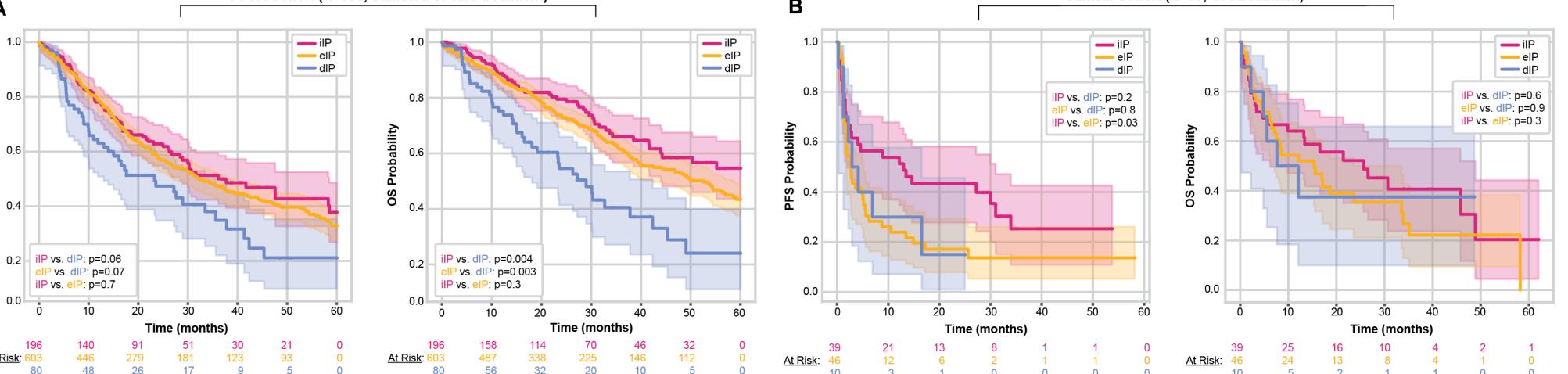
- Lymphocyte density in cancer epithelium and fraction of hot cancer epithelial patches were significantly associated with PFS (HR=0.64, q=0.04 and HR=0.69, q=0.04, respectively; Table 2).
- Notably, in PD-L1 (-) patients (N=43, TPS \leq 1%), iIP patients (orange line) had longer PFS than eIP and dIP patients (blue line; HR=0.35, p=0.02; Fig. 6B, C). No difference in PFS was observed for PD-L1 (+) patients (N=43, TPS >1%).

Figure 3. Distribution of TME-related features and immune phenotypes in NSCLC cohorts.



In the TCGA and clinical cohorts, lymphocyte density was extracted after PathExplore deployment in the cancer epithelium (A) and cancer stroma (B). C) IPs were predicted based on patch-level thresholds of hot patches in cancer and stroma in the TCGA and clinical cohorts.

Figure 4. Association of immune phenotype with PFS and OS.



Cox regression using predicted IPs was used to predict PFS and OS in A) the TCGA cohort and B) the clinical cohort, the latter of which consisted exclusively of CPI-treated patients.

Table 2. PFS regression results with covariates in clinical cohort. Features retaining significance after FDR correction are shown.

Feature	p	q	HR (95% CI)
Number of lymphocytes relative to all predicted cells in cancer epithelium	0.005	0.04	0.64 (0.46, 0.87)
Density of lymphocytes in cancer epithelium	0.006	0.04	0.64 (0.46, 0.88)
Percentage of "hot" patches in cancer epithelium	0.007	0.04	0.69 (0.53, 0.90)

Figure 5. Association of immune phenotype with PD-L1 TPS in the clinical cohort.

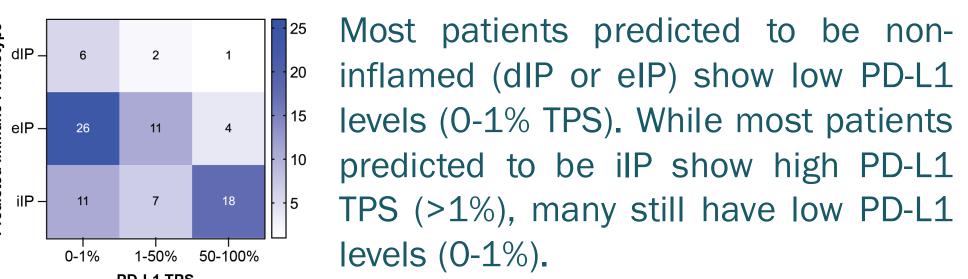
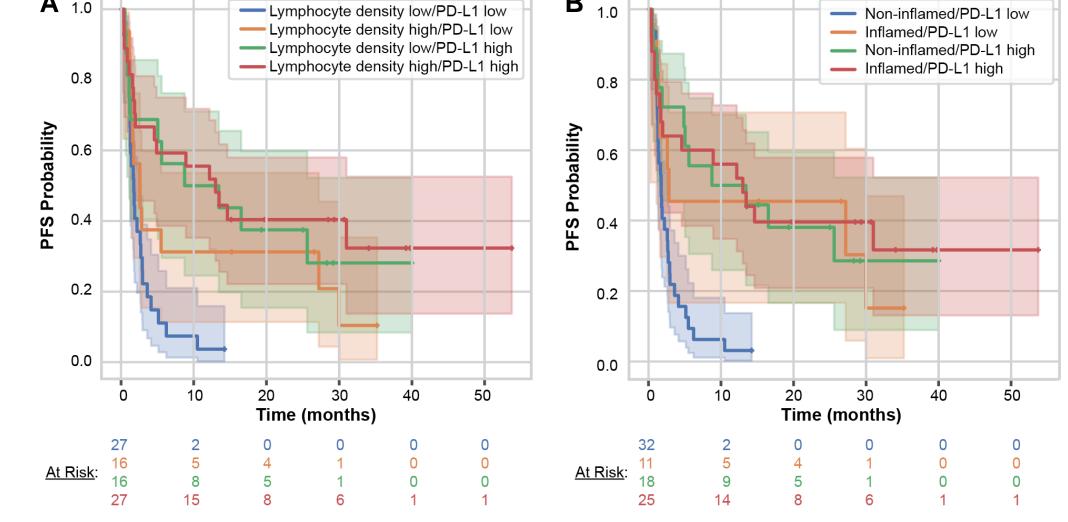


Figure 6. Immune inflamed phenotype associates with improved PFS in CPI-treated NSCLC patients independent of PD-L1 status



	Covariate	р	HR (95% CI)		
Inclusion of lymphocyte density in cancer epithelium as covariate	PD-L1 Low	0.002	2.42 (1.38, 4.23)		
	High TIL Density	0.06	0.62 (0.37, 1.03)		
	Prior treatment	0.53	0.93 (0.74, 1.17)		
	Histology	0.83	0.95 (0.57, 1.59)		
	Age	0.91	1.00 (0.97, 1.02)		
Inclusion of iIP prediction as covariate	PD-L1 Low	0.002	2.38 (1.36, 4.16)		
	iIP prediction	0.04	0.55 (0.32, 0.97)		
	Prior treatment	0.49	0.92 (0.74, 1.16)		
	Histology	0.64	0.88 (0.52, 1.49)		
	Age	0.92	1.00 (0.98, 1.03)		

Multivariable Cox regression using A) lymphocyte density binarized at the median cutoff or B) IP predictions as covariates was used to predict PFS in the clinical cohort. Lymphocyte density was binarized at the median value, while ilP patients were compared to non-inflamed (eIP and dIP). iIPinflamed status significantly correlates with better PFS (p=0.04). High lymphocyte density also correlates with better PFS but the effect does not reach statistical significance (p=0.06). C) Association between covariates and survival. Similar trends were observed for OS (data not shown).

We developed a data-driven approach for

CONCLUSIONS

- predicting IPs using patch-level lymphocyte densities in cancer epithelium and cancer stroma derived from H&E-stained samples.
- Model-predicted IPs associate with OS in the TCGA NSCLC dataset and with PFS in a CPItreated clinical NSCLC cohort. Association of IP and PFS was independent of PD-L1 status, potentially allowing the identification of PD-L1(-) patients who may derive greater benefit from CPI.

AFFILIATIONS

²Incendia Therapeutics, Boston, MA ³Explicyte, Bordeaux, France ⁴Institut Bergonié, Bordeaux, France *Contributed equally to study

publications@pathai.com

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* PathExplore is for research use only. Not for use in